

## "Education for Knowledge, Science and Culture" Shikshanmaharashi Dr. Bapuji Salunkhe



## Vivekanand College, Kolhapur (Autonomous) Department of Biotechnology (Optional)

#### B.Sc. Part I Semester I & II

Sr.	Semester	Paper	Name Of Paper	Total Marks
No.		Paper I	Basics of Biotechnology I	100
1	Semester I	Paper II	Basics of Biotechnology II	100
2		Paper III	Microbiology	100
	Semester II	Paper IV	Basics Cell IV biology and genetics	100

CBCS Syllabus to be implemented from September 2021 onwards

"Education for Knowledge, Science and Culture" -Shikshanmaharashi Dr. Bapuji Salunkhe

# Shri Swami Vivekanand Shikshan Sanstha's VIVEKANAND COLLEGE, KOLHAPUR (AUTONOMOUS) Department of Biotechnology Optional

## Academic year 2021-22 B.Sc. I Biotechnology Optional

#### COS for Semester I And II

Semester	Course outcome		
Semester I			
Paper I	DSC-1009A Paper I Basics of Biotechnology I		
	1 to the course students Will able to		
	and the province proteins with their structural level.		
	CO 2: Classify types of vitamins & able to state their deficiency		
	syndromes.		
	CO 3: Specify types of Diabetes, causes, and remedies.		
	CO 4: Qualitative and quantitative analysis of biomolecules		
Paper II	DSC-1009A Paper-II Basics of Biotechnology II		
	At the end of this course students will able to;		
	CO 1: Isolate & purify particular protein.		
	CO2. Explain the principle of centrifugation.		
	CO 3. I Inderstand the working of Microscope.		
	CO 4: Discuss the instrumentation & working of UV visible		
	spectroscopy		
	•		
Semester II	DSC-1009 B Paper -III Microbiology		
Paper -III	At the end of this course students will able to;		
1 mp	CO 1: Elucidate the harmful activities of bacteria.		
	CO 2: Design media to culture specific bacterial strain.		
	CO 3: Conclude importance of sterilization		
	CO 4: Compare various types of staining.		
Paper -IV	DSC-1009 B Basics of Cell biology and genetics		
	At the end of this course, students will able to;		
	CO 1: List various cell organelles with functions.		
	CO 2: Differentiate Prokaryotic & Eukaryotic cells.		
	CO 3: Elaborate the Mendelian Genetics.		
	CO 4: Predict how crossing over helps in species diversity &		
	evolution		



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HEAD
DEPARTMENT OF BIOTECHNOLOGY (OPTIONAL)
VIVEKANAND COLLEGE, KOLHAPUR
(//L10/HOM/JMGUS)

#### VivekanandCollege(Autonomous), Kolhapur

CHOICE BASED CREDIT SYSTEM SYLLABUS

## ForBachelor of SciencePart - I BIOTECHNOLOGY (Optional)

#### Introduction

This syllabus is framed to give sound knowledge with understanding of Biotechnology to undergraduate students at first year of three years of B.Sc. degree course. Students learn Biotechnology as a separate subject from B.Sc. I. The goal of the syllabus is to make the study of Biotechnology popular, interesting and encouraging to the students for higher studies including research.

The new and updated syllabus is based on a basic and applied approach with vigor and depth. At the same time precaution is taken to make the syllabus comparable to the syllabi of other universities and the needs of industries and research.

The syllabus is prepared after discussion at length with number of faculty membersofthe subject and experts from industries and research fields. The unitsof the syllabus are well defined, taking into consideration the level and capacity of students.

#### Objectives:-

- 1) To make the students knowledgeable with respect to the subject and its practicableApplicability.Due to which student become familiar withdifferent techniques in biotechnology at under graduatelevel
- 2) To promote understanding of basic and advanced concepts inBiotechnology.
- 3) To expose the students to various emerging areas ofBiotechnology,(Medical biotechnology)
- 4) To prepare students for further studies, helping in their bright career in the subject.
- 5) To expose the students to different processes used in industries and inresearch field.
- 6) To prepare the students to accept the challenges in lifesciences.
- 7) To develop skills required in various industries, research labs and in the fieldof human health.



	Semester I- Paper I- Basics of Biotechnology I	
	UNIT I	Lectures (30)
1	Biotechnology: definition, history, Branches, scope Amino acids and Protein:-Introduction, General structure of amino acids, Structural classification of amino acids based on R side chain, single letter code, Reaction of amino acids, Structure of peptide bond, biological functions of protein, structural levels of protein- Primary, Secondary, Tertiary (Myoglobin), Quaternary (Hemoglobin)  Vitamins- Introduction, Types, Roles, Deficiency	15
2	UNIT II	
<b>~</b>	Carbohydrate: :- General classification of carbohydrates, ring formation in monosaccharide, mutarotation, formation of glycosidic bond, study with respect to structure, chemical properties, hydrolysis of disaccharides (e.g. sucrose, maltose, lactose, ), oligosaccharides, polysaccharides (e.g. starch, glycogen, Cellulose) biological functions of carbohydrates.  Diabetes militias causes, type, remedies	
	Lipid: Definition, Fatty Acids, types, Classification of lipids- Simple lipid- (triacylglycerols & waxes) Compound lipid- (phospholipids, sphingolipids, cerebrosides, Lipoproteins), Derived lipid – e.g. cholesterol Properties of lipid. Functions of Lipids.	15



Paper II Basisc of Biotechnology II	
Unit I  Protein Purification: Method of cell disruption (Blenders, grinding with brasives, presses, enzymatic method, sonication); Salt participation-lating in, salting out, organic solvent precipitation, dialysis, ultra ditration.  Centrifugation- Basic principles, RCF, Sedimentation coefficient, wedberg's constant, Types of centrifuge: Desktop, High speed and Iltracentrifuge, Preparative centrifugation: Differential and density radient centrifugation  Unit II  Microscopy: a) General principles of microscopy- Image formation, magnification, numerical aperture, resolving power of microscope and rorking distance. b) Ray diagram, special features, applications and comparative study of compound microscope and Electron Microscope Scanning and Transmission Electron Microscope).  V-visible spectroscopy and Colorimetry-  Introduction to spectroscopy, properties of electromagnetic radiation (UV and Visible range, Electromagnetic spectrum, Electronic Transitions, minciple, Instrumentation with respect to colorimeter and single beam proceed to the principle of Sterilization: Methods of sterilization  Physical agents: i) temperature-dry heat, moist heat ii) Radiation  V, Gamma radiation iii) filters- membrane filter.  Chemical agents:- Phenol & Phenolic compounds, Alcohol,	
<b>Protein Purification</b> : Method of cell disruption (Blenders, grinding with abrasives, presses, enzymatic method, sonication); Salt participation-Salting in, salting out, organic solvent precipitation, dialysis, ultra filtration.	15
Centrifugation- Basic principles, RCF, Sedimentation coefficient, Svedberg's constant, Types of centrifuge: Desktop, High speed and Ultracentrifuge, Preparative centrifugation: Differential and density gradient centrifugation	
Unit II	
Microscopy: a) General principles of microscopy- Image formation, magnification, numerical aperture, resolving power of microscope and working distance. b) Ray diagram, special features, applications and comparative study of compound microscope and Electron Microscope (Scanning and Transmission Electron Microscope).  UV-visible spectroscopy and Colorimetry-	
Introduction to spectroscopy, properties of electromagnetic radiation (UV and Visible range, Electromagnetic spectrum, Electronic Transitions,	
Principle, Instrumentation with respect to colorimeter and single beam spectrophotometer. Principle, Instrumentation, Applications of UV and Visible spectrophotometer and colorimeter Lambert-Beer's law,	15
<ul> <li>A. Concept of Sterilization: - Methods of sterilization</li> <li>a)Physical agents: i) temperature-dry heat, moist heat ii) Radiation</li> <li>U.V, Gamma radiation iii) filters- membrane filter.</li> <li>b) Chemical agents: - Phenol &amp; Phenolic compounds, Alcohol,</li> <li>Heavy metals (e.g. mercury).</li> <li>c) Gaseous agents- Ethylene oxide, formaldehyde</li> </ul>	

#### Reference Books :-

- 1. Text book of biotechnology- Pradip parihar student ed. Jodpur (2004)
- 2. Biotechnology expanding horizons- B. D. Singh, Kalyani Publishe
- 3. Elements of biotechnology- P. K. Gupta, Rastogi publications.
- 4. Biotechnology- V. Kumarsan, Saras publication.
- 5. A text book of biological chemistry- M. S. Yadav, Dominant publishers.
- 6.Outline of biochemistry- Conn & Stumph
- 7. Principles of Biochemistry- Jeffory, Zubey
- 8. Biochemistry-Lubert Stryer
- 9. Textbook of Biotechnology R. C. Dubey. 10)Biochemistry by Lehninger
- 10. Biochemistry U. Satyanarayana
- 11.Biochemistry Glick & Pasterneck
- 12.Practical Biochemistry principles and techniques Willson and Walker
- 13. Protein purification by Robert Scope
- 14. Biophysical chemistry- Nath Upadhyay



Semester II- Paper III-Microbiology	
 Unit I	Lectures(30
History of Microbiology: - Contributions of Anton van	
Leeuwenhoek, Alexander Fleming, Louis Pasteur, Robert Koch, Joseph Lister.	
Introduction to types of Microorganisms – Bacteria, Algae,	
Fungi, Protozoa and Viruses.	
Beneficial and harmful activities of microorganisms, Applied	
Branches of Microbiology	15
Morphology and cytology of Bacteria	
A. Morphology of Bacteria – i) Size, ii) Shape, iii) Arrangements	
B. Cytology of Bacteria – Structure of Typical Bacterial Cell	
a) Structure and functions of Bacterial cell parts	
i) Cell wall ii) Cell membrane iii) Capsule and slime layer iv)Flagella v)	
Pilli vi) Nuclear material vii) Mesosome viii)Ribosome	
Unit II	
A.Culture media and pure culture techniques: Common components of media and their functions Peptone, Yeast extract, NaCl, Agar and Sugar	
<b>B.</b> Culture media - a) Living Media (Lab. animals, plants, bacteria, embryonated eggs, tissue cultures), b) Non living media – i)Natural, ii) Synthetic, iii) Semi synthetic, iv)Differential, v) Enriched, vi) Enrichment, vii) Selective.	
C. Methods for isolation of pure cultures- Streak plate, pour plate, spread plate.	15
D. Stains and staining procedures -	
a. Definition of dye and stain	
b. Classification of stains – Acidic, Basic and Neutral	
d .Principle, Procedure, Mechanism and application of staining	
procedures	
i) Simple staining	
ii) Negative staining	
iii) Differential staining: Gram staining and Acid fast staining	
m) Billoronida saming. Grain staining and Mola last staining	



	Paper II-Basics in Cell biology & Genetics	
	Unit I	Lectures(30)
1	History of Cell biology:- Introduction of cell and concept of prokaryotic and Eukaryotic cell. Cell biology before 19th century, cell biology in19th century, formulation of cell theory, protoplasm theory, germplasm theory, cell biology in 20th century- organismal theory, Branches of Cell Biology.  Structure and function of Cell organelles- ultra structure and function of cell membrane, golgibodies, Endoplasmic reticulum (rough & smooth ) Ribosome, cytoskeleton structure( actin, microtubules),Mitochondria, chloroplast, Lysosomes, peroxisomes, Nucleus. Cell division and cell cycle- phases of cell cycle, Mitosis.Meosis	15
	UNIT II	
	Mendels law of Inheritance – principal of segregation, Independent assortment, Dominance, Mendelian genetics in humans.  Varity of gene expression – modifiers, suppressors, pleiotropic gene, multiple allele. Interaction of gene- Epstasis, complimentary gene, duplicate gene.  Linkage definition, coupling and repulsion hypothesis, linkage groups. Crossing over –Mechanism and theory. Structural and numerical changes in chromosomes. Extra chromosomal inheritance-mitochondrial and plastids.	15

#### Reference Books-

- 1. Fundamentals of Microbiology- Frobisher
- 2. Microbiology-Pelczar.
- 3. General Microbiology- Stanier.
- 4. Text book of Microbiology- Ananthnarayan & Panikar.





- 5. Cell and molecular biology- Arumugham
- 6.Cell and molecular biology- De Robertis
- 7. Cytology genetics and evolution- Agarwal and Varma
- 8.Cell biology- C. B. Pawar
- 9.Cell-Cooper.
- 10.Cell biology- Gilard Karp
- 11. Biology of Microorganisms- Brock
- 12. Cell biology Albert Brown



#### Practical syllabus

## (Practical Examination to be conducted annually)

### Lab.Exercises in Basics of Biotechnology Credit -1

	Title	Hr. Allotted
SN		Allotted
1	Preparation of Molar and Normal solutions	
	Molar solution of Sucrose.	
	Normal solutions	
	Molar solution of alkali-NaOH and Acid HCL	
	Preparation of Buffers	
2	Isolation of casein from milk	
3	Study of Lamberts Beer's Law by copper sulphate method.	
4	Estimation of glucose by DNSA method (Graphical)	
5	Isolation of starch from potato	
6	Determination of acid value of given fat	
7	Estimation of DNA by Diphenylamine method (by calculation)	
8	Estimation of RNA by Orcinol method (By calculation)	
9	Estimation of reducing sugar by Benedict's method	
10	Identification of given amino acid by paper chromatography	
11	Estimation of protein by Biuret method (Calculation)	
	Lab Exercises in cell biology and Microbiology	
1	Use, care and study of Compound Microscope	
2	Preparation of Culture media	
	Pentone water Nutrient broth and Nutrient Agar	
	-MacConkey's Agar ,Sabroud's Agar Starch Agar ,Milk Agar	
3	Microscopic Examination of Bacteria	
	1. Monochrome staining	
	2. Negative Staining	
	3. Gram's Staining	
	Hanging drop technique- Motility.	
4	Isolation, colony characters, Gram's staining and motility of	
	Bacteria isolated from-	
	- Air-( solid impaction technique)	

Water- (dilution and spreading plate technique)	
Enumeration of Bacteria from soil by total viable court P	
- should all of some lab equipments: Autoclary II-4 -: 0	
TAI, CEIIIIIII OF Colorimeter Water bath of 1	
Counter, Water distillation unit	
Sums of Medellin genetics	
Study of mitosis	
Isolation of chloroplast	
Study of effect of organic solvent and temperature members and	
permeability permeability	
	Water- (dilution and spreading plate technique.) Enumeration of Bacteria from soil by total viable count- Pour plate technique. Mounting and identification of mould- Penicillium, Aspergillus Demonstration of some lab equipments:- Autoclave, Hot air Oven, Incubator, LAF, Centrifuge, Colorimeter, Water bath, Colony Counter, Water distillation unit Sums of Medellin genetics Study of mitosis Isolation of chloroplast Study of effect of organic solvent and temperature membrane permeability

#### Books recommended for Practical

- 1) Stains and Staining procedures by Desai and Desai.
- 2) Introduction to Practical Biochemistry by D. Plummer, J Wiley and Sons.
- 3) Bacteriological techniques by F. J. Baker.
- 4) Introduction to Microbial techniques by Gunasekaran.
- 5) Biochemical methods by Sadashivan and D. Manickam.
- 6) Laboratory methods in Biochemistry by J. Jayaraman.
- 7) Experimental Microbiology Patel & Patel



### List of minimum equipments-

- 1) Hot air oven 1
- 2) Incubator 1
- 3) Autoclave 1
- 4) Refrigerator 1
- 5) Medical microscopes 10 nos. for one batch
- 6) Digital weighing balance 1
- 7) Digital pH meter 1
- 8) Centrifuge 1
- 9) Colorimeter 1
- 10) Distilled Water Plant 1
- 11) Laminar air flow cabinet 1
- 12) Colony counter 1
- 13) Water bath 1
- 14) Arrangements for gas supply and fitting of two burners per table.
- 15) One working table of 6' x 2½' for two students.
- 16) One separate sterilization room attach to the laboratory (10' x 15')
- 17) At least one wash basin for a group of five students
- 18) One separate instrument room attached to lab (10' x 15')
- 19) One laboratory for one batch including working tables (6' x  $2\frac{1}{2}$ ') per two students for one batch 20) Store room (10' x 15')



#### **Practical Examination**

- (A) The practical examination will be conducted on two consecutive days for three hours per day per batch.
- (B) Each candidate must produce a certificate from the Head of the Department in her/his college, stating that he/she has completed satisfactory practical course online laid down from Academic Council on the recommendations ofBoard of Studies and that the journal has been properly maintained. Every candidatemust have recorded his/her observations in the laboratory journal and have written areport on each exercise performed. Every journal is to be checked and signed periodically by a member of teaching staff and certified by the Head of the Departmentat the end of the year. Candidates must produce their journals at the time of practical examinations.

Note:- At least 80% Practical should be covered in practical examination.



#### Nature of Question paper and distribution of marks for Practical

#### Examination

Q.1 One major practical(Biochemistry)	10M
Q.2 One Miner Practical (Biochemistry)	05 M
Q.3 One major practical (Cell biology and Microbiology)	10M
Q.4One Miner Practical (Cell biology and Microbiology)	05 M
Q.5 Spotting	10M
O.6 Journal	10M



#### SCHEME OF MARKING FOR (THEORY)

Sem	Core	Marks	Evaluation	Sections	Answer	Standard of
	Course				Books	passing
1	DSC-	80	Semester	Two sections each of	As per	35%
	9A		wise	40 marks	instruction	(28 marks)
2	DSC-	80	Semester	Two sections each of	As per	35%
	9B		wise	40 marks	instruction	(28 marks)

#### SHEME OF MARKING (CIE) Continues Internal Evaluation

Sem	Core	Marks	Evaluation	Sections	Answer	Standard of
	Course				Books	passing
1	DSC-9A	20	Semester	One	As per	35%
			wise		instruction	(7marks)
2	DSC-9B	20	Semester	One	As per	35%
		7				
			wise		instruction	(7marks)

#### SHEME OF MARKING (PRACTICAL)

Sem	Course	Marks	Evaluation	Section	Standard of
					•
					passing
I & II	DSC 9A &	50	Annual	As per	35%
	DSC 9B			instruction	(18marks)

<sup>\*</sup>A separate passing is mandatory

